The effect of aliskiren on the renal dysfunction following unilateral ureteral obstruction in the rat

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Received March 16, 2016; Accepted April 12, 2016; Epub August 5, 2016; Published August 15, 2016

Abstract: Purpose: To investigate the effect of blocking renin-angiotensin system by direct renin inhibition using aliskiren on the renal dysfunction following reversible unilateral ureteral obstruction (UO). Methods: Wistar rats underwent reversible left UO for 72 hours. Group-Alsk (n=12) received aliskiren (30 mg/kg/day) dissolved in water starting one day before creating UO and continued until the terminal experiment five days post reversal when renal functions were measured using clearance techniques. Group-Vx (n=12) underwent similar protocol but had water only. Gene expression analysis of some markers of kidney injury was measured using PCR technique. Results: In Group-Vx, renal blood flow (RBF) and glomerular filtration rate (GFR) in the left kidney were significantly lower than the right kidney (1.82±0.12 vs. 3.19±0.40, P=0.001 and 0.81±0.08 vs. 1.44±0.09, P=0.004, respectively). However, left fractional excretion of sodium (FE$_{Na}$) was higher than the right FE$_{Na}$ (0.80±0.15 vs. 0.55±0.04, P=0.05). Comparing the left obstructed kidney in Group-Alsk vs. Group-Vx, RBF and GFR were higher in Group-Alsk (2.44±0.30 vs. 1.82±0.12, P=0.049 and 1.02±0.11 vs. 0.81±0.08, P=0.07, respectively). The left renal FE$_{Na}$ was lower in Group-Alsk but did not reach statistical significance (0.54±0.07 vs. 0.80±0.15, P=0.07). Aliskiren also decreased the gene expressions of NGAL, KIM-1 and p53. Conclusion: Direct renin inhibition by aliskiren appears to have protective effect on the renal dysfunction and on the markers of renal injury following UO indicating a potential clinical benefit of this agent. Further, this data and the previous studies indicate that blocking renin-angiotensin system at any level has a protective effect in obstructive nephropathy.

Keywords: Ureteral obstruction, aliskiren, renal function

Introduction

Ureteral obstruction (UO) is a relatively common clinical problem and usually cause pain and if prolonged might lead to renal impairment [1]. The renal damage is due to the interaction of various intra-renal systems leading to alterations in hemodynamic and tubular renal functions [2]. Renin-angiotensin system (RAS) plays a central role in orchestrating these alterations and the blockade of RAS at different levels has been shown to ameliorate these obstruction-induced changes [3-6].

The RAS consists of several peptides which interact in a cascade fashion to produce angiotensin-II. This cascade can be blocked at different levels by agents such as angiotensin converting enzyme (ACE) inhibitors and angiotensin receptor blockers (ARBs). Similarly, this blockade can be achieved by direct renin inhibition. Since renin catalyzes the first and rate-limiting step in the RAS cascade, direct renin inhibition is expected, at least theoretically, to be more effective than ACE inhibitors or ARBs in blocking the RAS. Moreover, ACE inhibitors and ARBs have been shown to result in a compensatory increase in renin release which might dampen the effects of these agents [7, 8] as demonstrated by the larger effect of renin inhibitor enalkiren in achieving renal vasodilation compared to the ACE inhibitor captopril [9, 10].

Historically, direct renin inhibition has not been used successfully in the clinical setup until recently due to the lack of potency or good bioavailability. Aliskiren is the first oral direct renin inhibitor which has been approved by the US Food and Drug Administration. Aliskiren has been shown to have beneficial effects in differ-
ent renal conditions [11-13]. Few studies have investigated the role of direct renin inhibition by aliskiren on the renal dysfunction induced by UO [14-18]. For instance, aliskiren has been recently shown to ameliorate the renal inflammatory process induced by UO [14, 16]. Aliskiren also reduced renal fibrosis in a mice [14] and rat [15, 17] models of unilateral UO. However, the effect of direct renin inhibition by aliskiren on the alterations in renal hemodynamic and tubular functional parameters following UO has not been investigated yet. Thus, the aim of this study was to investigate this effect in a rat model of reversible unilateral UO.

Materials and methods

Studies were performed in male Wistar rats weighing 211-261 g at the time of UO. Rats were housed in standard cages and kept in a 12-hour light-dark cycle at 20°C. They were fed a standard rat chow and had free access to water. Animals were fasted for 12 hours before the experimental procedures but had water ad libitum. The experimental protocol was approved by the local animal research ethics committee.

Ureteral obstruction and reversal

The following procedures were carried out under aseptic conditions. Rats were placed on a heated table to maintain rectal temperature at 37-38°C. Each animal was anesthetized with ketamine hydrochloride (80 mg/kg, intraperitoneally, Pantex Holland B.V., Holland) and xylazine Hydrochloride (8 mg/kg, intraperitoneally, Troy Laboratory PTY Limited, NSW, Australia). The left ureter was exposed via a midline abdominal incision and obstructed by placing a 3 mm length of bisected PVC tubing (0.58 mm internal diameter) around the left midureter as described previously [19, 20]. The ureter was then occluded by constricting the tubing with a 4-0 silk suture. At the end, the wound was closed in layers.

The reversal of UO was performed 72 hours later using similar anesthesia as described above. Using a dissecting microscope, and through the same incision, the obstructing tube was identified and removed. Full release of the obstruction was confirmed by observation of a free flow of urine across the site of obstruction. The wound was then closed in layers.

Aliskiren/vehicle administration

Aliskiren (Novartis Pharma Stein AG, Stein, Basle, Switzerland) was dissolved in 0.5 ml of water (vehicle) and administered by gavage immediately after preparation as single daily dose of 30 mg/kg. Control animals received only 0.5 ml of water. In the two groups, treatment was commenced 24 hours before UO and continued daily throughout the period of obstruction and for 5 days thereafter until the time of the terminal experiment.

Experimental groups

Animals were divided into two groups:

Group-Vx (n=12): Rats which underwent left UO for 72 hours and received water only.

Group-Alsk (n=12): Rats which underwent left UO and received aliskiren.

Surgical procedure in the terminal experiment

All rats underwent terminal experiment five days following reversal of UO. Animals were anaesthetised with pentobarbital sodium (60-70 mg/kg, intraperitoneally; Sigma Life Science, St Louis, USA) and the trachea was cannulated. Following cannulation of a femoral vein with polyethylene tubing (PE-50), anaesthesia was maintained by a continuous infusion of pentobarbital sodium (12.5 mg/kg/hr) and a sustaining infusion of 0.9% saline was established at a rate of 50 µl/min using an infusion pump. A femoral artery was cannulated with similar tubing used in the femoral vein and the tip of the cannula was positioned just below the level of the left renal artery. The cannula was connected to a pressure transducer (Memscap, Skoppum, Norway). The blood pressure signal was amplified using a bridge Amp (ADInstruments, Castle Hill, Australia), digitised using Power Lab 4/30 and Lab Chart version-6 software (ADInstruments, Australia) and displayed on a computer screen. The arterial cannula was also used to obtain blood samples throughout the procedure as required. Both kidneys were exposed through a midline abdominal incision and the upper ureters were cannulated with polyethylene tubing (PE-10) for the collection of urine into pre-weighed micro-capped tubes. The urine volume was determined gravimetrically.
On completion of surgery, the sustaining infusion of 0.9% saline was replaced by one composed of Fluorescein isothiocyanate-inulin (FITC-inulin, Sigma-Aldrich, St Louis, USA) (2.5 mg/ml) and para-aminohippuric acid (PAH, Sigma-Aldrich, St Louis, USA) (0.2% w/v) in 0.9% saline. A priming dose of 2 ml of the same solution was infused over 2 minutes. Animals were allowed 2 hours to equilibrate before being subjected to the experimental protocol.

**Experimental protocol and assays**

The terminal experiment was performed to determine the hemodynamic functions including glomerular filtration rate (GFR) and renal blood flow (RBF) as well as the tubular functions (urine volume (UV), urinary sodium (U\textsubscript{Na}V) and fractional excretion of sodium (FE\textsubscript{Na})). The experimental protocol consisted of two 30-minute clearance periods. Arterial blood samples (0.4 ml) taken at the beginning and end of the clearance periods were immediately centrifuged. Plasma samples (125 µl) were frozen to be assayed later. The plasma was replaced by an equal volume of saline and the erythrocytes were re-suspended by gentle vortexing and returned to the animal. The hematocrit was determined. Finally, after euthanizing the animals, the kidneys were removed, weighed and prepared for gene expression analysis (vide infra).

Urine and plasma samples were assayed for sodium level using a flame photometer (Corning, Halstead, Essex, England). GFR was estimated from the clearance of inulin. RBF was calculated using the formula [RBF=ERPF/(1-hematocrit)], where the PAH clearance was used to estimate ERPF (effective renal plasma flow). The values of GFR, RBF, UV, U\textsubscript{Na}V and FE\textsubscript{Na} were calculated as the average of the two clearance periods and were corrected for kidney weight.

**Gene expression analysis**

The middle part of each kidney was excised, immediately snap-frozen in liquid nitrogen and stored at -80°C for a later measurement of gene expression of neutrophil gelatinase-associated lipocalin (NGAL) and kidney injury molecule-1 (KIM1) which were used as markers of renal tubular injury and the apoptotic protein p53 which was used as a marker of apoptotic response.

Total RNA was extracted using TRI Reagent® Solution (Life Technologies Corporation, NY, USA) according to the manufacturer protocol. Quality and quantity of the extracted RNA was estimated using NanoDrop instrument (Thermo Fisher Scientific Inc., DE, USA). First-strand cDNAs were prepared in duplicates from 2.0 µg of the extracted RNA with GoScript™ Reverse Transcriptase (Promega Corporation, Wisconsin, USA) in the presence of RNasin® Plus RNase inhibitor (Promega Corporation, Wisconsin, USA) according to manufacturer protocols. Prepared cDNA was used as a template for the relative gene expression analysis by real-time PCR using TaqMan® chemistry on QuantStudio™ 7 Flex Real-Time PCR system (Applied Biosystems, CA, USA). The reaction mixture consisted of 75 ng cDNA, TaqMan® Universal Master Mix (Applied Biosystems, CA, USA), 0.6 µM of forward and reverse primers and 0.25 µM of the fluorescent probes (Biosearch Technologies, Inc., CA, USA). Sequences of primers and fluorogenic probes are listed in Table 1. Primers and probes were designed.

### Table 1. Forward and reverse primers and fluorogenic probe sequences used for real time quantitative PCR analysis

<table>
<thead>
<tr>
<th>Gene</th>
<th>Gene Bank Reference</th>
<th>5’-3’ Sequence</th>
<th>Size</th>
</tr>
</thead>
<tbody>
<tr>
<td>NGAL</td>
<td>NM_130741.1</td>
<td>Forward CTGTTCCCACCCGACCAATGC</td>
<td>69</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Reverse CCACCCACATCCCCGTCGA</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Probe d FAM-TGACAATGACAGACCCAGC-BHQ-1</td>
<td></td>
</tr>
<tr>
<td>kIM-1</td>
<td>NM_173149.2</td>
<td>Forward GCGCTGGAATAATCACACTGTAAG</td>
<td>75</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Reverse GCAACGGAACGCACACATAG</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Probe d FAM-TCCCTTTGAGGAAGCCCGAAG-BHQ-1</td>
<td></td>
</tr>
<tr>
<td>p53</td>
<td>NM_030989.3</td>
<td>Forward CGAGATGTTGCCGAGAAGCTGAATG</td>
<td>86</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Reverse GTCTCGGAGTAGGATGAGT</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Probe d FAM-CCTTGGATTAAGGATGCCCAGTGC-BHQ-1</td>
<td></td>
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using the online RealTimeDesign™ software (Biosearch Technologies, Inc., CA, USA) in a way that at least one of the primers was spanning an exon-exon junction within their respective gene. Peptidylprolyl isomerase A (ppia) (cyclophilin A) was used as the endogenous control gene for normalization between samples and it was multiplexed within each PCR reaction.

Calculated CT values were used to estimate changes in gene expression of target genes using delta-delta CT formula.

The results are expressed as the mean fold change of gene expression in the left obstructed kidney compared to right control kidney in the respective group.

Statistical analysis

Statistical analysis was performed using SPSS V16.0. Results were expressed as means ± SEM. One-way factorial ANOVA was used for comparison of variables between the two groups and between the control and obstructed kidneys within each group. P value of less than 0.05 was considered statistically significant.

Results

The mean arterial blood pressure and heart rate in Group-Vx and Group-Alsk were similar (121±3 vs. 119±4, P=0.8 and 461±7 vs. 463±17, P=0.5, respectively).

Glomerular and tubular functions

In Group-Vx which had left UO but did not receive aliskiren, left RBF, five days following
reversal of UO, was 57% of the right RBF (1.82±0.12 vs. 3.19±0.40, P=0.005). Similarly, left GFR was 55% that of the right GFR (0.81±0.08 vs. 1.44±0.09, P=0.0001) (Figure 1). With the decrease in both RBF and GFR, there was an increase in the FE\textsubscript{Na} in the left kidney compared to the right kidney (0.80±0.15 vs. 0.55±0.04, P=0.05). This was associated with a decrease in both the UV and U\textsubscript{Na}V in the left kidney but both did not also reach statistical significance (8.4±1.6 vs. 15.5±4.7 and 1.1±0.3 vs. 2.1±0.6, respectively, P=0.08 for both) (Figure 2).

In Group-Alsk which received aliskiren, the left RBF was 71% of the right RBF (2.44±0.34 vs. 3.42±0.45, P=0.09) and the left renal GFR was 72% of the right GFR (1.02±0.11 vs. 1.41±0.14, P=0.04) (Figure 1). As shown in Figure 2, the UV, U\textsubscript{Na}V and FE\textsubscript{Na} of the left kidney were not significantly different from those of the right kidney (11.0±2.9 vs. 17.3±4.1 (P=0.2), 1.09±0.26 vs. 2.16±0.47 (P=0.07) and 0.54±0.07 vs. 0.59±0.07 (P=0.6).

When Group-Alsk was compared to Group-Vx, all variables in the right kidneys in both groups were similar (P>0.05 for all variables). However, when the left obstructed kidneys in the two groups were compared, the left RBF was higher in Group-Alsk (2.44±0.34 vs. 1.82±0.12, P=0.05). The left GFR was also higher in Group-Alsk but did not reach statistical significance (1.02±0.11 vs. 0.81±0.08, P=0.07) (Figure 1). As shown in Figure 2, left renal FE\textsubscript{Na} was lower in Group-Alsk but also did not reach statistical significance (0.54±0.07 vs. 0.80±0.15, P=0.07) (Figure 2). However, the UV and U\textsubscript{Na}V were similar in both groups (P>0.05 for both variables). All variables in the right kidneys in both groups were similar (P>0.05 for all variables).

Gene expression analysis results

As demonstrated in Figure 3, in Group-Vx, there was 5.5±1.3 fold increase in the expression of NGAL in the left obstructed kidney compared to the right control kidney, whereas in Group-Alsk, there was only 2.7±0.26 fold increase (P=0.03). Similarly, the left to right kidney expression of KIM-1 was lower in Group-Alsk but this did not reach statistical significance (115±46 vs. 52±9, P=0.09). The left to right kidney expression of p53 was also significantly lower in the Group-Alsk (1.34±0.05 vs. 1.10±0.02, P=0.01).

Discussion

In the current study, we have demonstrated for the first time that direct renin inhibition by aliskiren prior to, during and following reversal of unilateral UO has resulted in a significant improvement in the hemodynamic and tubular renal functional parameters as well as an attenuation of the gene expression of some of the markers of renal injury.

RAS plays an important role in the development of UO-induced renal dysfunction. Angiotensin-II is synthetized de novo in the kidney and the level of intrarenal angiotensin-II has been shown to increase in the obstructed kidney [4,
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21]. This increase appears to mediate the decrease of GFR and RBF following UO as shown by the ability of both ACE inhibitors or ARBs to partially prevent the reductions in GFR and RBF observed in UO [3]. Further, ARBs increased the level of several important sodium transporters [22], indicating a potential role of RAS in the renal tubular dysfunction observed in obstructed kidneys. These effects might not be merely due to decreased production of angiotensin-II but also could be due to the associated decrease in the level of several other substances and cytokines which were shown to play an important role in the UO-induced renal impairment [23, 24]. Collectively, these data and the data from the current study indicate that blocking RAS at any level results in an improvement of the UO-induced renal dysfunction.

Whether direct renin inhibition by aliskiren is more superior to ACE inhibitors or ARBs in attenuating the UO-induced renal alterations would require a direct comparative study using the three types of medications in the same model. However, it is expected that the effect might depend on the drug dose, species and model of obstruction including the period, laterality and reversibility of obstruction. In general, direct renin inhibition might be more effective than other methods of blocking RAS because it does not only decreases angiotensin-II production but also decreases plasma renin activity, an effect which is not observed in ACE inhibitors or ARBs [7-10]. Regardless of the superiority of any agent, blocking RAS at more than one level has been shown to be more beneficial in preventing the renal alterations following UO [17].

In the current study, aliskiren improved the RBF and to some extent the GFR in the previously obstructed kidney. These effects are similar to the effect of ACE inhibitors or ARBs as shown previously [3]. In this study, there was some improvement in the GFR although it did not reach statistical significance. The exact reason is difficult to ascertain but it could be due to the possible selective effect of angiotensin-II on efferent as compared to afferent glomerular arterioles. Angiotensin-II was shown to have preferential effect on the efferent arterioles which results in the maintaining of GFR especially in some pathological condition [25]. Therefore, decreasing the level of angiotensin-II by aliskiren might have resulted in eliminating this selective effect on efferent arterioles and hence a lower effect on GFR compared to its effect on RBF.

The current data indicates that aliskiren does not only affect the hemodynamic functions but also the tubular renal functions. So aliskiren significantly reduced the increase in FE_{Na} observed in the obstructed kidney and hence its ability to concentrate urine. This is consistent with the previous findings of the effect of RAS blockade by ACE inhibitors or ARBs [22, 26]. The lack of significant effect of aliskiren on total UV and U_{Na}V is probably due to the fact that these parameters are not determined only by the FE_{Na} which has decreased by aliskiren but also by GFR which has increased somewhat by aliskiren resulting in a lack of significant change in these two parameters as a net result.

Aliskiren has been shown to have a higher specificity for human and mouse renin compared to rat renin [27]. In this study, the dose of aliskiren was similar to that used in other studies in rat models [11]. The significant effect of aliskiren on hemodynamic and tubular parameters indicates that bioavailability was not a major issue in this study.

In addition to ameliorating the alterations in renal functional parameters, aliskiren decreased the expression of some of the markers of renal tubular injury such as NGAL and KIM-1. Each of the markers is heavily expressed by different parts of the renal tubules and hence, it was possible to determine the exact site of aliskiren effect. KIM-1 has been shown to be strongly expressed and released by injured proximal tubular epithelial cells [28] whereas NGAL is synthesized in the thick ascending limb of Henle’s loop and collecting ducts [29]. In this study, the effect of aliskiren administration on NGAL was more pronounced than the effect on KIM-1. Although, it is difficult to determine the reason for this differential effect from this study, the results might indicate a weaker effect of aliskiren on the proximal tubules where the majority of the filtered sodium gets reabsorbed. This is consistent with the weak effect of aliskiren on the FE_{Na} demonstrated in the current study.

In addition to its effect on NGAL and KIM-1, aliskiren decreased the expression of p53 which is a marker of apoptotic response. Renal
p53 expression has been shown previously to increase following ureteric obstruction [23] and the mechanism of this rise remains unclear, although increased renin production, ischemia, and elevated intra-tubular pressure has been suggested to be relevant [30]. The results of this study lend support to the role of RAS in this rise. Regardless of the exact mechanism, the observed effect on p53 in the current study indicates a protective effect on the ongoing renal fibrosis observed in obstructive nephropathy [17, 23] and is consistent with the previous findings [14].

The obstruction model used in this study is similar to the clinical scenario of a transiently obstructing ureteral calculus. The beneficial effect of aliskiren on the kidney might be of clinical interest in patients with obstructing ureteral calculus especially those with a single kidney or those with compromised renal functions; however, further clinical studies are required to extrapolate the results to humans.

In conclusion, direct renin inhibition by aliskiren before, during and after release of unilateral UO appears to have a protective effect on the hemodynamic and tubular renal functional alterations as well as on the markers of renal injury. This indicates a potential benefit of this agent in the clinical set-up. Further, this data and the previous findings indicate that blocking RAS at any level has a protective effect in obstructive nephropathy.

Acknowledgements

This study was funded by an individual grant from the College of Medicine & Health Sciences, United Arab Emirates University.

Disclosure of conflict of interest

None.

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