

## Original Article

# Modulatory influence of *Parkia biglobosa* protein isolate on testosterone and biomarkers of oxidative stress in brain and testes of streptozotocin-induced diabetic male rats

Bolajoko Idiat Ogunyinka<sup>1</sup>, Babatunji Emmanuel Oyinloye<sup>1,2</sup>, Foluso Oluwagbemiga Osunsanmi<sup>1</sup>, Andrew Rowland Opoku<sup>1</sup>, Abidemi Paul Kappo<sup>1</sup>

<sup>1</sup>Biotechnology and Structural Biology (BSB) Group, Department of Biochemistry and Microbiology, University of Zululand, Kwadlangezwa 3886, South Africa; <sup>2</sup>Department of Biochemistry, College of Sciences, Afe Babalola University, PMB 5454, Ado-Ekiti 360001, Nigeria

Received February 11, 2016; Accepted August 19, 2016; Epub September 30, 2016; Published October 15, 2016

**Abstract:** *Parkia biglobosa* seed an important household spice commonly consumed in Nigeria is believed to possess antioxidant activity that may exert modulatory effects in diabetes and diabetic complications. This study investigated the modulatory potential of *Parkia biglobosa* protein isolate (PBPI) on serum testosterone (sTT) level as well as its influence on biomarkers of oxidative stress in brain and testes of streptozotocin-induced diabetic male rats. Animals were made diabetic by single intraperitoneal administration of streptozotocin (STZ; 60 mg/kg body weight). PBPI (200 or 400 mg/kg body weight) was given orally by gavage or insulin (5 U/kg, i.p.) was administered daily to STZ-induced diabetic rats for 28 days. The results revealed a significant elevation in thiobarbituric acid reactive substances (TBARS) levels in the brain and testes of diabetic rats. This was closely associated with a concomitant reduction in levels of sTT and reduced testes weight, a noticeable decline in the glutathione-S-transferase (GST), superoxide dismutase (SOD) and catalase (CAT) as well as total glutathione (Total GSH) level in the brain and testes of diabetic rats. Interestingly, treatment with PBPI efficiently prevented the alterations witnessed in the serum sTT and also ameliorated various alterations in the biomarkers of oxidative stress (TBARS, Total GSH, GST, SOD and CAT) in brain and testes of diabetic rats. These results provide evidence that PBPI could protect the brain and testicular tissues against oxidative stress induced by STZ, via modulation of serum testosterone concentration and also by enhancing antioxidant defence system in STZ-diabetic rats.

**Keywords:** Antioxidant, brain, oxidative stress, *Parkia biglobosa*, STZ-induced diabetes, testosterone

## Introduction

Diabetes mellitus (DM) has been identified as the fourth leading cause of death globally, it ranks high among the common non-communicable chronic metabolic disorder affecting people in both developed and developing countries in recent times [1-3]. The exponential increase in the prevalence of DM has been linked to obesity and increasing sedentary behaviours, urbanization, modernization, genetics and family history as well as nutritional imbalance associated with consumption of high energy, fat and cholesterol rich diets among others [4, 5]. This insidious metabolic disease is characterized by chronic hyperglycaemia (increased blood glu-

cose level) together with impaired carbohydrate, protein and lipid metabolism due to deficiency in insulin secretion or resistance to insulin action or both [6, 7].

Recent documented evidence suggests that prolonged hyperglycaemia promotes the upregulation and overproduction of reactive oxygen species in the mitochondrial, which ultimately leads to oxidative stress [8-10]. This is believed to occur via well established mechanisms, including activation of polyol pathway, increased intracellular formation of advanced glycation end-products, activation of protein kinase C, increased production of superoxide radicals by the mitochondrial electron transport chain and

## Modulatory influence of *P. biglobosa* on brain and testes in STZ-diabetic rats

a general decline in endogenous antioxidant capacity [11-13]. To a great extent, experimental evidence reveals that hyperglycaemia-induced oxidative stress enhances the progression of diabetes and its associated complications [10, 14].

Oxidative stress is known to arise when there is an imbalance between free radical production (especially reactive oxygen species; ROS) and endogenous antioxidant defense system. This shift in balance is associated with oxidative damage to a wide range of biomolecules including lipids, proteins, and nucleic acids, which may eventually impair normal functions of various tissues and organs [15, 16]. Diabetic cerebral disorders have been established at a neurochemical, electrophysiological, structural and cognitive level. Although diabetes is not often considered to have damaging effects on the brain, accumulating evidence in literature reveals that long-term diabetes results in a variety of delicate cerebral disorders, which occur more frequently than is normally believed, even though the pathogenesis is poorly understood [17]. Recurring episodes of hypoglycaemic and hyperglycaemic in diabetes may have profound influence on brain chemistry, particularly on brain neurotransmitters and associated enzymes [18]. Excessive ROS production from the auto-oxidation of elevated intracellular glucose levels has been reported as one of the mechanisms underlying diabetic neuronal injury [19]. It has been established that elevated ROS levels can make the brain susceptible to oxidative stress and damage due to the brain's high oxygen demand as well as its abundant lipid concentration and relative poor antioxidant defense system compared to other tissues [20-22].

Similarly, there is increasing evidence that male reproductive dysfunction in experimental animal model and human with diabetes is one of the consequences of the imbalance between free radical production and antioxidant defense system associated with diabetic conditions [23-27]. Diabetes may affect male reproductive functions in various ways including decreased sperm count and motility, decreased testicular testosterone production, testicular morphological changes and reduced libido [27-31]. The role of medicinal plants in assuaging the devastating effects of hyperglycaemia-induced oxida-

tive stress associated with diabetes in various tissues and organs have been recognized.

*Parkia biglobosa* popularly known as 'African Locust Beans' is widely consumed in Nigeria and many other West African countries as a spice for flavouring foods [4]. Both aqueous and methanolic extracts of *P. biglobosa* have been reported to possess some beneficial and pharmacological potential such as hepatoprotective, hypoglycaemic, hypolipidaemic, antimicrobial and anti-inflammatory activities in experimental animals [4, 32, 33]. To the best of our knowledge, no study has been carried out to explore the modulatory influence of *Parkia biglobosa* protein isolate (PBPI) on serum testosterone concentration as well as biomarkers of oxidative stress in brain and testes of STZ-induced diabetic rats. We demonstrate here that PBPI at a dose of 200 or 400 mg/kg bw exhibited a significant reversal effect in all biochemical parameters (notably in serum testosterone; sTT and the extent of lipid peroxidation measured by thiobarbituric acid reactive substances; TBARS) measured in serum and tissues (brain and testes) of STZ-challenged rats. Thus, in treatment of neurotoxicity and testicular toxicity, PBPI confer protection against oxidative stress induced by STZ.

### Materials and methods

#### *Plant material*

Raw and fermented *Parkia biglobosa* seeds were obtained from a local market in Ijebu-Ode, Ogun state, Nigeria. Import permit (P0060156) from the Department of Agriculture, Forestry and Fisheries (DAFF; Republic of South Africa) was obtained. The raw seeds sample were identified and authenticated by the Chief botanist in the Department of Botany, University of Zululand and a voucher specimen (B07) was deposited in the University Herbarium.

#### *Animals*

Male Sprague-Dowley rats, weighing about 250-290, were obtained from the animal house of the Department of Biochemistry and Microbiology, University of Zululand. The animals were kept for acclimatization for 7 days prior to the commencement of the study; they were maintained at standard conditions of temperature and relative humidity, with a 12-hour

## Modulatory influence of *P. biglobosa* on brain and testes in STZ-diabetic rats

light/dark cycle. The animals were provided with standard rat pellets and water *ad libitum*. The animal experimental protocols were in accordance with the recommendations of the institutional animal ethical committee (UZREC 171110-030-RA level O2 Dept 2014/74).

### Chemicals

All the chemicals used this study were procured from Sigma Chemical Co. (St. Louis, MO, USA), Merck (South Africa) and Sciencell Research Laboratories (Carlsbad, CA, USA).

### Preparation of seed extract of *Parkia biglobosa*

Oven-dried (50°C) seeds of fermented *Parkia biglobosa* were pulverized using an electric blender to obtain a coarse powder. One kilogram of uniform powdered seeds was defatted with hexane to obtain the defatted extract. The defatted extract was air dried and then extracted (1:10 w/v) with butanol to remove possible anti-nutrients. Protein isolate was obtained from the defatted extract using the method described by Nkosi et al. [34]. Briefly, the dry defatted extract was re-suspended in distilled water at pH 10. The resultant suspension was filtered to remove debris and the filtrate adjusted to pH 5, followed by centrifugation at 5000 rpm for 15 minutes at 4°C. The supernatant was discarded while the pellet containing the protein isolate was retained and freeze-dried to yield a brown extract. The lyophilized extract was kept dry until needed.

### Induction of diabetes

Following an overnight fasting, diabetes was induced in the selected rats by a single intraperitoneal injection of freshly prepared STZ (Sigma-Aldrich Co.) at a dose of 60 mg/kg body weight; dissolved in 0.1M, pH 4.5 ice cold citrate buffer [35]. Diabetes was confirmed in rats 72 hours after STZ administration by measuring the fasting blood glucose (FBG) levels from blood samples collected from rat-tail vein using a glucometer (Accu Check Roche, Germany). Rats with FBG level above 300 mg/dl were considered diabetic and selected for the study. Treatment commenced on the fourth day and continued for a period of twenty-eight days.

### Experimental design

Seventy healthy, male rats (averaging 12 weeks old) were divided into seven groups of ten ani-

mals each and treated as follows: Group 1 (control) was given citrate buffer only. Group 2 (PI 200ND), non-diabetic rat, was given citrate buffer and protein isolate (200 mg/kg body weight). Group 3 (PI 400ND), non-diabetic rat, was given citrate buffer and protein isolate (400 mg/kg body weight). STZ-induced diabetic rats were divided in four groups (Groups 4-7). Group 4 (STZ), diabetic control. Group 5 (STZ I), positive control, was given insulin (5 U/kg, i.p.). Group 6 (STZ PI 200) was diabetic and received protein isolate (200 mg/kg body weight). Group 7 (STZ PI 400) diabetic rats that received protein isolate (400 mg/kg body weight). PBPI treatments were given orally by gavage for 28 days.

### Collection and processing of blood and tissue samples

At the end of the 28 days of treatment, the rats were fasted overnight and then sacrificed under anesthesia; blood samples were obtained by cardiac puncture in plain tubes without anticoagulants, left for 1 hour to coagulate then centrifuged at 3000 rpm for 15 min at 4°C to obtain serum. The whole brain and testes were collected, washed in saline, blotted dry and weighed. Portions of the rats' brain and testes were homogenized in 56 mM Tris-HCl buffer (pH 7.4) containing 1.15% KCl, and then centrifuged at 10,000 x g for 15 minutes to obtain the supernatants that were stored at -80°C until needed for analysis.

### Biochemical and enzyme estimation

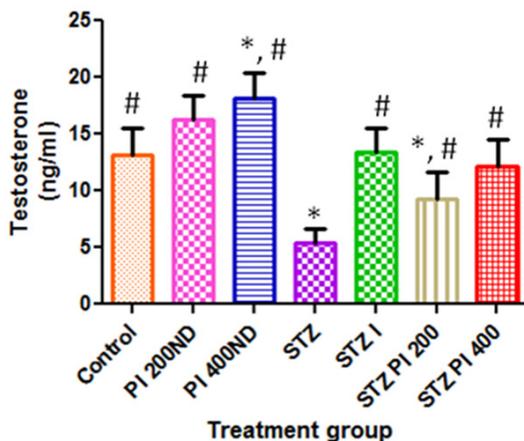
The extent of lipid peroxidation in the brain and testes homogenate was determined spectrophotometrically by measuring the formation of thiobarbituric acid reactive substances (TBARS) according to the method described by Varshney and Kale [36], and this was expressed in terms of malondialdehyde (MDA), which is the end product of the reaction. Serum testosterone level was estimated using ELISA kit (Cayman Ltd., USA) according to the manufacturer's instructions while the brain and testes level of total glutathione (Total GSH) as well as the activities of glutathione-S-transferases (GST), superoxide dismutase (SOD) and catalase (CAT), were determined using the corresponding assay kits (Sciencell Research Laboratories, Carlsbad, USA) according to manufacturer's instructions.

## Modulatory influence of *P. biglobosa* on brain and testes in STZ-diabetic rats

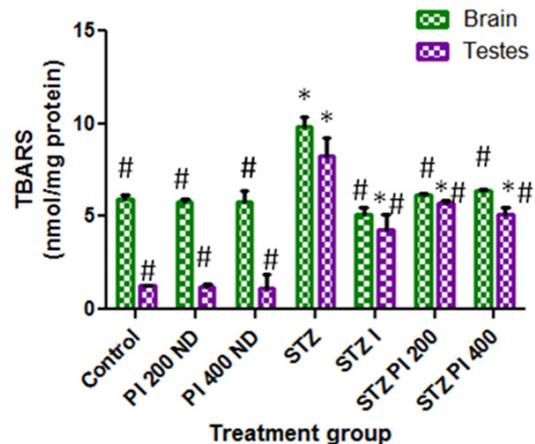
**Table 1.** Effects of PBPI on testes weight of STZ-induced diabetic rats

Testes	Control	PI 200ND	PI 400ND	STZ	STZ I	STZ PI 200	STZ PI 400
Weight (g)	3.29 ± 0.10	3.57 ± 0.25	3.61 ± 0.10	2.60 ± 0.55	3.42 ± 0.08	3.34 ± 0.11	3.40 ± 0.55

Group 1: Control; given citrate buffer only, Group 2: PI 200ND; non-diabetic rat, given citrate buffer + protein isolate (200 mg/kg body weight), Group 3: PI 400ND; non-diabetic rat, given citrate buffer + protein isolate (400 mg/kg body weight), Group 4: STZ; untreated diabetic rats, Group 5: STZ I; diabetic rats treated with insulin (5 U/kg, i.p.), Group 6: STZ PI 200; diabetic rats treated with protein isolate (200 mg/kg body weight). Group 7: STZ PI 400; diabetic rats treated with protein isolate (400 mg/kg body weight).



**Figure 1.** Effects of PBPI on serum testosterone level of STZ-induced diabetic rats. Testosterone (ng/ml). Data are presented as mean ± S.D. (n = 10). Mean differences are significant (P < 0.05) when compared with: \*control group, #STZ only.



**Figure 2.** Effects of PBPI on lipid peroxidation levels in brain and testes of STZ-induced diabetic rats. TBARS (nmol/mg protein). Data are presented as mean ± S.D. (n = 10). Mean differences are significant (P < 0.05) when compared with: \*control group, #STZ only.

### Statistical analysis

All data are presented as the mean ± standard deviation. The data were analyzed by one-way ANOVA followed by Dunnett's test for multiple comparisons using Graphpad Prism (GraphPad Prism version 5.0, GraphPad Software Inc., San Diego, CA). The differences were considered significant at P < 0.05.

### Results

#### Effects of PBPI on testes weight and serum testosterone level of STZ-induced diabetic rats

**Table 1** and **Figure 1** show the testes weight and serum testosterone level in the experimental rats, respectively. STZ significantly decreased the weight of the testes and serum testosterone level in diabetic animals compared to control rats. Treatment with PBPI (200 or 400 mg/kg bw) for 28 consecutive days caused a significant increase in the testes weight and serum testosterone level as compared to diabetic control.

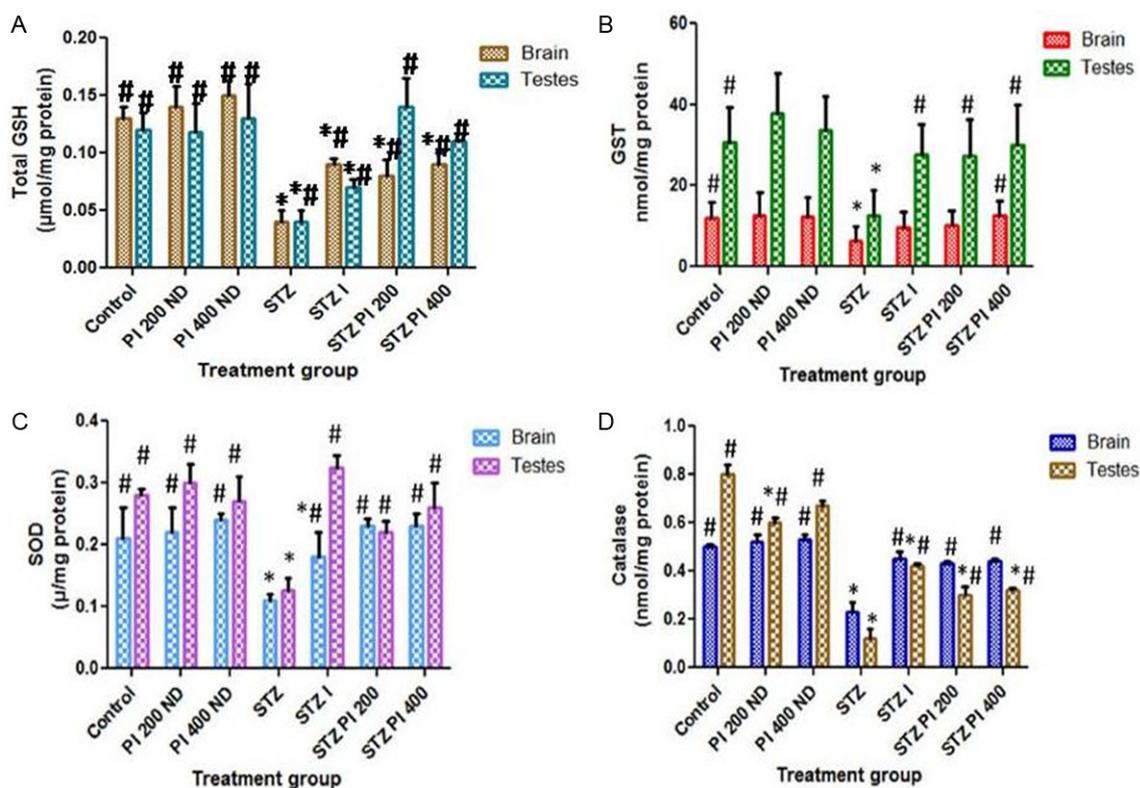
#### Effects of PBPI on lipid peroxidation levels in brain and testes of STZ-induced diabetic rats

The extent of lipid peroxidation in brain and testes of experimental rats are shown in **Figure 2**. Induction of diabetes led to a significant increased level of TBARS in brain and testes of diabetic rats. Interestingly, the increased level of TBARS in the brain was reversed near normal when PBPI (200 or 400 mg/kg bw) was administered. In like manner, PBPI (200 or 400 mg/kg bw) supplementation in the treatment groups considerably decreased the TBARS level in the testes of diabetic rats.

#### Effects of PBPI on antioxidant parameters in brain and testes of STZ-induced diabetic rats

**Figure 3** represents the various alterations in the antioxidant status of experimental rats. Induction of diabetes was accompanied by a significant decline in the level of total GSH as well as in the activities of GST, SOD and CAT in the brain and testes of diabetic rats compared

## Modulatory influence of *P. biglobosa* on brain and testes in STZ-diabetic rats



**Figure 3.** Effects of PBPi on antioxidant parameters in brain and testes of STZ-induced diabetic rats. (A) Total GSH ( $\mu\text{mol}/\text{mg}$  protein), (B) GST ( $\text{nmol}/\text{mg}$  protein), (C) SOD ( $\mu/\text{mg}$  protein), (D) CAT ( $\text{nmol}/\text{mg}$  protein). Data are presented as mean  $\pm$  S.D. ( $n = 10$ ). Mean differences are significant ( $P < 0.05$ ) when compared with: \*control group, #STZ only.

to the control. These altered antioxidant status were restored significantly after the administration of PBPi (200 and 400 mg/kg bw).

### Discussion

Brain cell injury or impairment as well as reproductive dysfunction are common complications of diabetes, which may occur via several mechanisms [9, 37]. It appears that diabetes, which is characterized by hyperglycaemia-induced oxidative stress, results in multi-organ failure and this account for most common causes of death in people with diabetes [38, 39]. The present work is one of the series of studies showing relationship between STZ-induced free radicals damage and the potentials of nutraceuticals/medicinal plants in reducing oxidative stress while concurrently improving various tissues and organ functions affected by free radicals damage. The results presented in this study demonstrated that PBPi (200 or 400 mg/kg bw) significantly ameliorated all indices of alterations in all biochemical parameters evalu-

ated in serum and tissue homogenates (brain and testes) of STZ-challenged rats. This complemented previously identified protective roles of *P. biglobosa* in human health and disease [4, 33].

Reduction in testes weight and serum testosterone level was observed in diabetic rats. This is in agreement with similar studies reported in literature [40, 41]. Generally, testicular function is controlled via two independent coordinated functions; the biosynthesis of androgens by Leydig cells and the production of spermatozoa in the epithelium of seminiferous tubules. The reduction in testosterone level in this study may arise from oxidative damage in the testes, which may be due to a decrease in the function of both Leydig cells (testosterone producing cell) and Sertoli (supporting cell), which might be due to a reduction in insulin secretion [26, 42].

In the measurement of toxicity in STZ-induced diabetes, enhanced lipid peroxidation plays a

## Modulatory influence of *P. biglobosa* on brain and testes in STZ-diabetic rats

significant role and also serves as one of the most important manifestations of oxidative stress and damage in diabetic complications [43, 44]. Earlier work and sufficient evidence have demonstrated that neuronal cells are sensitive to oxidative insults due to the fact that the brain is susceptible to oxidative stress and damage as a result of its high oxygen demand, abundant lipid content, and relative poor antioxidant defense mechanisms as compared to other tissues [19-21]. Increased TBARS level in brain and testes homogenates in this study was an indication of damage and dysfunction in these organs. This finding is consistent with earlier reports [37, 44-46]. ROS-induced cellular damage due to chronic hyperglycaemia occurs via lipid peroxidation of unsaturated fatty acids, which alters cellular function [47]. The reversal in the elevated levels of TBARS particularly in the brain suggests the protective influence of PBPI against oxidative stress in STZ-induced diabetes.

In addition to the increased level of lipid peroxidation witnessed in the STZ diabetic animals, the endogenous antioxidant defence mechanisms (notably, Total GSH, GST, SOD and CAT) were also altered in the brain and testes. Uncontrolled hyperglycaemia, which promotes free radical generation, may be the underlying causative factor [48]. Our observation is also in agreement with previous documented reports [38, 49-51]. It is interesting to note that in this particular experiment, PBPI rescued total GSH for both brain and testes whereas it did not have any pronounced effect on brain GST activity but did rescue testes GST activity. The possible reason for the observed effect may be as a result of the overwhelming effect of the ROS generated on the antioxidant systems in the brain particularly, GST. GPx and GST work in harmony with GSH and decompose decomposing superoxide and hydrogen peroxide before interacting to form the more reactive hydroxyl radical. It is widely accepted that antioxidant enzymes provide first line defence against ROS in response to oxidative challenges [52]. Enzymatic antioxidants play very important role in protecting the cell against the overwhelming deleterious effect of ROS in hyperglycaemia-induced oxidative stress [53].

SOD and CAT are the two important radical scavenging enzymes. SOD catalyzes the conversion of superoxide radical to  $H_2O_2$ , a less

reactive intermediate, thereby protecting the cell from the deleterious effect of superoxide radicals while CAT is required to neutralize this reactive specie ( $H_2O_2$ ) to water ( $H_2O$ ) and molecular oxygen ( $O_2$ ) [54-56]. On the other hand, glutathione defends the cells from oxidative damage by reducing disulfide bonds of cytoplasmic proteins to cysteines. The glutathione system includes glutathione, glutathione reductase, glutathione peroxidases and glutathione-S-transferases [57]. The decreased activities of these antioxidants in diabetic conditions might result in reduced protection against free radical damage. This is an indication that PBPI may possess bioactive constituents that are capable of scavenging free radicals, preventing their potential induction of cellular damage.

In conclusion, the observations from this study confirms that STZ (60 mg/kg bw) has an adverse effect on serum testosterone level and on the antioxidant status in the brain and testes of diabetic rats via induction of lipid peroxidation in our animal model. Thus, our study proposes that PBPI showed a modulatory effect by attenuating the above lipid peroxidation in STZ-induced diabetic rats in a dose-dependent manner, with 400 mg/kg bw identified as having particular efficacy with no evidence of toxicity. Taken together, these results provide evidence that PBPI could protect the brain and testicular tissues against oxidative stress induced by STZ, via modulation of serum testosterone concentration and also by enhancing antioxidant defence system in STZ-diabetic rats. Therefore, characterization of the protein isolate from *P. biglobosa* should be further studied in order to elucidate the active component and the exact mechanism by which PBPI interferes with STZ toxicity in animal models.

### Acknowledgements

Research reported in this article was supported by the South African Medical Research Council (SAMRC) through funding received from the South African National Treasury. Its contents are solely the responsibility of the authors and do not necessarily represent the official views of the South African Medical Research Council. More so, the support of the University of Zululand Research Committee (S1335/14) towards this study is greatly appreciated. The authors will also like to acknowledge Mr. Mathula Lance Ngwenya for technical expertise regarding experimental animals.

# Modulatory influence of *P. biglobosa* on brain and testes in STZ-diabetic rats

## Disclosure of conflict of interest

None.

**Address correspondence to:** Dr. Abidemi Paul Kappo, Biotechnology and Structural Biology (BSB) Group, Department of Biochemistry and Microbiology, University of Zululand, KwaDlangezwa 3886, South Africa. E-mail: KappoA@unizulu.ac.za

## References

- [1] Wu Y, Ding Y, Tanaka Y and Zhang W. Risk factors contributing to type 2 diabetes and recent advances in the treatment and prevention. *Int J Med Sci* 2014; 11: 1185-1200.
- [2] Shah A and Afzal M. Prevalence of diabetes and hypertension and association with various risk factors among different Muslim populations of Manipur, India. *J Diabetes Metab Disord* 2013; 12: 52.
- [3] Meaney B. Diabetic foot care: prevention is better than cure. *J Ren Care* 2012; 38: 90-98.
- [4] Ogunyinka BI, Oyinloye BE, Adenowo AF and Kappo AP. Potentials of some plant-derived foods in the management of diabetes and associated Complications. *Afr J Tradit Complement Altern Med* 2015; 12: 12-20.
- [5] Ramachandran A, Snehalatha C, Shetty AS and Nanditha A. Trends in prevalence of diabetes in Asian countries. *World J Diabetes* 2012; 3: 110-117.
- [6] Schmatz R, Perreira LB, Stefanello N, Mazzanti C, Spanevello R, Gutierrez J, Bagatini M, Martins CC, Abdalla FH, da Silva Serres JD and Zanini D. Effects of resveratrol on biomarkers of oxidative stress and on the activity of delta aminolevulinic acid dehydratase in liver and kidney of streptozotocin-induced diabetic rats. *Biochimie* 2012; 94: 374-383.
- [7] Abdel-Sattar EA, Elberry AA, Harraz FM, Ghareib SA, Nagy AA and Gabr SA. Antihyperglycemic and hypolipidaemic effects of the methanolic extract of Saudi mistletoe (*Viscum schimperi* Engl). *J Adv Res* 2011; 2: 171-177.
- [8] Tangvarasittichai S. Oxidative stress, insulin resistance, dyslipidemia and type 2 diabetes mellitus. *World J Diabetes* 2015; 6: 456-480.
- [9] Matough FA, Budin SB, Hamid ZA, Alwahaibi N and Mohamed J. The role of oxidative stress and antioxidants in diabetic complications. *Sultan Qaboos Univ Med J* 2012; 12: 5-18.
- [10] Giacco F and Brownlee M. Oxidative stress and diabetic complications. *Circ Res* 2010; 107: 1058-1070.
- [11] Tiwari BK, Pandey KB, Abidi AB and Rizvi SI. Markers of oxidative stress during diabetes mellitus. *J Biomarkers* 2013; 2013: 378790.
- [12] Campos C. Chronic hyperglycemia and glucose toxicity: pathology and clinical sequelae. *Postgrad Med* 2012; 124: 90-97.
- [13] Rains JL and Jain SK. Oxidative stress, insulin signaling, and diabetes. *Free Radic Biol Med* 2011; 50: 567-575.
- [14] Shailey S and Basir SF. Strengthening of antioxidant defense by *Azadirachta indica* in alloxan-diabetic rat tissues. *J Ayurveda Integr Med* 2012; 3: 130-135.
- [15] Rahman T, Hosen I, Islam MT and Shekhar HU. Oxidative stress and human health. *Adv Biosci Biotechnol* 2012; 3: 997-1019.
- [16] Lobo V, Patil A, Phatak A and Chandra N. Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacogn Rev* 2010; 4: 118-126.
- [17] Biessels GJ, Kappelle AC, Bravenboer B, Erkelens DW and Gispen WH. Cerebral function in diabetes mellitus. *Diabetologia*. 1994; 37: 643-650.
- [18] Mohamed MI, Aly MS, Abdel T and Mahmoud MS. Effect of panax ginseng on the activity of cholinesterase in different tissues of experimentally induced diabetes in rats. *J Egypt Soc Toxicol* 2007; 37: 95-106.
- [19] El-Akabawy G and El-Kholy W. Neuroprotective effect of ginger in the brain of streptozotocin-induced diabetic rats. *Ann Anat* 2014; 196: 119-128.
- [20] Muriach M, Flores-Bellver M, Romero FJ and Barcia JM. Diabetes and the brain: oxidative stress, inflammation, and autophagy. *Oxid Med Cell Longev* 2014; 2014: 102158.
- [21] Patil R, Dhawale K, Gound H and Gadakh R. Protective effect of leaves of *Murraya koenigii* on reserpine-induced orofacial dyskinesia. *Iran J Pharm Res* 2012; 11: 635-641.
- [22] Rahman K. Studies on free radicals, antioxidants, and co-factors. *Clin Interv Aging* 2007; 2: 219-236.
- [23] Li M, Liu Z, Zhuan L, Wang T, Guo S, Wang S, Liu J and Ye Z. Effects of apocynin on oxidative stress and expression of apoptosis-related genes in testes of diabetic rats. *Mol Med Rep* 2013; 7: 47-52.
- [24] Roy S, Rahaman N, Ahmed F, Metya S and Sannigrahi S. Naringenin attenuates testicular damage, germ cell death and oxidative stress in streptozotocin induced diabetic rats: naringenin prevents diabetic rat testicular damage. *J Appl Biomed* 2013; 11: 195-208.
- [25] Ramalho-Santos J, Amaral S, Oliveira PJ. Diabetes and the impairment of reproductive function: possible role of mitochondria and reactive oxygen species. *Curr Diabetes Rev* 2008; 4: 46-54.
- [26] Alves MG, Martins AD, Rato L, Moreira PI, Socorro S, Oliveira PF. Molecular mechanisms

## Modulatory influence of *P. biglobosa* on brain and testes in STZ-diabetic rats

- beyond glucose transport in diabetes-related male infertility. *Biochimica et Biophysica Acta* 2013; 1832: 626-635.
- [27] Ballester J, Muñoz MC, Domínguez J, Rigau T, Guinovart JJ and Rodríguez-Gil JE. Insulin-dependent diabetes affects testicular function by FSH-and LH-linked mechanisms. *J Androl* 2004; 25: 706-719.
- [28] Jain GC and Jangir RN. Modulation of diabetes-mellitus-induced male reproductive dysfunctions in experimental animal models with medicinal plants. *Pharmacogn Rev* 2014; 8: 113-121.
- [29] Jangir RN, Jain GC. Diabetes mellitus induced impairment of male reproductive functions: a review. *Curr Diabetes Rev* 2014; 10: 147-57.
- [30] Glenn DR, McClure N, Lewis SE. The hidden impact of diabetes on male sexual dysfunction and fertility. *Hum Fert* 2003; 6: 174-179.
- [31] Ricci G, Catizone A, Esposito R, Pisanti FA, Vietri MT and Galdieri M. Diabetic rat testes: morphological and functional alterations. *Andrologia* 2009; 41: 361-368.
- [32] Adi K, Metowogo K, Mouzou A, Lawson-Evi P, Eklu-Gadegbeku K, Agbonon A, Lamboni C, Essien K, Aklidikou K and Gbeassor M. Evaluation of cardioprotective effects of *Parkia biglobosa* (jacq. benth) Mimosaceae stem bark. *J App Pharm Sci* 2013; 3: 60-64.
- [33] Odetola AA, Akinloye O, Egunjobi C, Adekunle WA and Ayoola AO. Possible antidiabetic and antihyperlipidaemic effect of fermented *Parkia biglobosa* (JACQ) extract in alloxan-induced diabetic rats. *Clin Exp Pharmacol Physiol* 2006; 33: 808-812.
- [34] Nkosi CZ, Opoku AR and Terblanche SE. Effect of pumpkin seed (*Cucurbita pepo*) protein isolate on the activity levels of certain plasma enzymes in CCl<sub>4</sub>-induced liver injury in low-protein fed rats. *Phytother Res* 2005; 19: 341-345.
- [35] Hule AK, Shah AS, Gambhire MN and Juvekar AR. An evaluation of the antidiabetic effects of *Elaeocarpus ganitrus* in experimental animals. *Indian J Pharmacol* 2011; 43: 56-59.
- [36] Varshney R and Kale RK. Effect of calmodulin antagonists on radiation induced lipid peroxidation in microsomes. *Int J Radiat Biol* 1990; 58: 733-743.
- [37] Ramzy MM, El-Sheikh AA, Kamel MY, Abdelwahab SA and Morsy MA. Mechanism of testicular protection of carvedilol in streptozotocin-induced diabetic rats. *Indian J Pharmacol* 2014; 46: 161-165.
- [38] Singh R, Bhardwaj P and Sharma P. Antioxidant and toxicological evaluation of *Cassia sopherain* streptozotocin-induced diabetic Wistar rats. *Pharmacognosy Res* 2013; 5: 225-232.
- [39] Cade WT. Diabetes-related microvascular and macrovascular diseases in the physical therapy setting. *Phys Ther* 2008; 88: 1322-1335.
- [40] Long L, Wang J, Lu X, Xu Y, Zheng S, Luo C and Li Y. Protective effects of Scutellarin on Type II diabetes mellitus-induced testicular damages related to reactive oxygen species/Bcl-2/Bax and reactive oxygen species/microcirculation/starving Pathway in Diabetic Rat. *J Diabetes Res* 2015; 2015: 252530
- [41] Khaki A, Khaki AA, Hajhosseini L, Golzar FS and Ainehchi N. The Anti-Oxidant Effects of Ginger and Cinnamon on spermatogenesis dysfunction of diabetes rats. *Afr J Tradit Complement Altern Med* 2014; 11: 1-8.
- [42] Ojewale AO, Olaniyan OT, Faduyile FA, Odukanmi OA, Oguntola JA and Dare BJ. Testiculo protective effects of ethanolic roots extract of *Pseudocedrela kotschyi* on alloxan induced testicular damage in diabetic rats. *Br J Med Med Res* 2014; 4: 548-563.
- [43] Subash-Babu P, Alshatwi AA and Ignacimuthu S. Beneficial antioxidative and antiperoxidative effect of cinnamaldehyde protect streptozotocin-induced pancreatic  $\beta$ -cells damage in wistar rats. *Biomol Ther* 2014; 22: 47-54.
- [44] Ramesh B, Karuna R, Sreenivasa RS, Haritha K, Sai MD, Sasis BRB and Saralakumari D. Effect of *Commiphora mukul* gum resin on hepatic marker enzymes, lipid peroxidation and antioxidants status in pancreas and heart of streptozotocin-induced diabetic rats. *Asian Pac J Trop Biomed* 2012; 2: 895-900.
- [45] Pari L and Murugan P. Tetrahydrocurcumin prevents brain lipid peroxidation in streptozotocin-induced diabetic rats. *J Med Food* 2007; 10: 323-329.
- [46] Nakhaee A, Bokaeian M, Akbarzadeh A, Hashemi M. Sodium tungstate attenuate oxidative stress in brain tissue of streptozotocin-induced diabetic rats. *Biol Trace Elem Res* 2010; 136: 221-31.
- [47] Florence NT, Benoit MZ, Jonas K, Alexandra T, Désiré DDP, Pierre K and Théophile D. Antidiabetic and antioxidant effects of *Annona muricata* (Annonaceae), aqueous extract on streptozotocin-induced diabetic rats. *J Ethnopharmacol* 2014; 151: 784-790.
- [48] Rolo AP and Palmeira CM. Diabetes and mitochondrial function: role of hyperglycemia and oxidative stress. *Toxicol Appl Pharmacol* 2006; 212: 167-178.
- [49] Aitken RJ and Roman SD. Antioxidant systems and oxidative stress in the testes. *Oxid Med Cell Longev* 2008; 1: 15-24.
- [50] Kumar G, Sharmila Banu G, Murugesan AG and Rajasekara Pandian M. Effects of *Helicteres isora* bark extracts on brain antioxidant status and lipid peroxidation in strepto-

## Modulatory influence of *P. biglobosa* on brain and testes in STZ-diabetic rats

- zotocin diabetic rats. *Pharm Biol* 2007; 45: 753-759.
- [51] Pari L and Latha M. Protective role of *Scoparia dulcis* plant extract on brain antioxidant status and lipid peroxidation in STZ diabetic male Wistar rats. *BMC Complement Altern Med* 2004; 4: 16.
- [52] Robertson RP and Harmon JS. Pancreatic islet  $\beta$ -cell and oxidative stress: The importance of glutathione peroxidase. *FEBS Letters* 2007; 581: 3743-3748.
- [53] Asmat U, Khan A and Khan MI. Diabetes mellitus and oxidative stress-a concise review. *Saudi Pharm J* 2015; 2015: doi: <http://dx.doi.org/10.1016/j.jsps.2015.03.013>.
- [54] Avelar TM, Storch AS, Castro LA, Azevedo GV, Ferraz L and Lopes PF. Oxidative stress in the pathophysiology of metabolic syndrome: which mechanisms are involved? *J Bras Patol Med Lab* 2015; 51: 231-239.
- [55] Al-Enazi MM. Protective effects of combined therapy of Rutin with Silymarin on experimentally-induced diabetic neuropathy in rats. *Pharmacol Phar* 2014; 5: 876-889.
- [56] Armagan A, Uz E, Yilmaz HR, Soyupek S, Oksay T and Ozcelik N. Effects of melatonin on lipid peroxidation and antioxidant enzymes in streptozotocin-induced diabetic rat testis. *Asian J Androl* 2006; 8: 595-600.
- [57] Liou GY and Storz P. Reactive oxygen species in cancer. *Free Radic Res* 2010; 44: 479-496.